

Moisture						
Our Reference:	UNITS	10350-1	10350-4	10350-8	10350-10	10350-12
Your Reference	-----	BH1	BH1	BH2	BH2	BH3
Depth	-----	0.1-0.3	3-3.45	0.5-0.95	3-3.45	0.1-0.4
Date Sampled		23/03/07	23/03/07	23/03/07	23/03/07	23/03/07
Type of sample		Soil	Soil	Soil	Soil	Soil
Moisture	%	8.4	11	12	12	7.1

Moisture						
Our Reference:	UNITS	10350-14	10350-15	10350-16	10350-17	10350-19
Your Reference	-----	BH3	BH4	BH4	BH5	BH5
Depth	-----	1.1-1.4	0.1-0.5	0.5-0.65	0.1-0.4	1.6-1.85
Date Sampled		23/03/07	23/03/07	23/03/07	24/03/07	24/03/07
Type of sample		Soil	Soil	Soil	Soil	Soil
Moisture	%	8.2	8.6	7.8	12	10

Moisture						
Our Reference:	UNITS	10350-22	10350-24	10350-25	10350-26	10350-27
Your Reference	-----	BH7	BH7	Dup3	Dup5	FB
Depth	-----	0.1-0.4	1.5-1.95	-	-	-
Date Sampled		24/03/07	24/03/07	23/03/07	26/03/07	23/03/07
Type of sample		Soil	Soil	Soil	Soil	Soil
Moisture	%	7.9	9.2	6.8	12	0.30

Asbestos ID - soils	UNITS	10350-1	10350-4	10350-8	10350-10	10350-12
Our Reference:	-----	BH1	BH1	BH2	BH2	BH3
Your Reference	-----	0.1-0.3	3-3.45	0.5-0.95	3-3.45	0.1-0.4
Depth		23/03/07	23/03/07	23/03/07	23/03/07	23/03/07
Date Sampled		Soil	Soil	Soil	Soil	Soil
Type of sample						
Sample Description	-	40g sand, rocks	40g sand, stone	40g sand, stone	40g sand, stone	40g sand, stone
Asbestos ID in soil	--	No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected
Trace Analysis	--	Respirable fibres not detected	Respirable fibres not detected	Respirable fibres not detected	Respirable fibres not detected	Respirable fibres not detected

Asbestos ID - soils	UNITS	10350-14	10350-15	10350-16	10350-17	10350-19
Our Reference:	-----	BH3	BH4	BH4	BH5	BH5
Your Reference	-----	1.1-1.4	0.1-0.5	0.5-0.65	0.1-0.4	1.6-1.85
Depth		23/03/07	23/03/07	23/03/07	24/03/07	24/03/07
Date Sampled		Soil	Soil	Soil	Soil	Soil
Type of sample						
Sample Description	-	40g sand, rocks	40g sand, rocks	40g sand, rocks	40g sand, rocks	40g sand, rocks
Asbestos ID in soil	--	No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected
Trace Analysis	--	Respirable fibres not detected	Respirable fibres not detected	Respirable fibres not detected	Respirable fibres not detected	Respirable fibres not detected

Asbestos ID - soils	UNITS	10350-22	10350-24
Our Reference:	-----	BH7	BH7
Your Reference	-----	0.1-0.4	1.5-1.95
Depth		24/03/07	24/03/07
Date Sampled		Soil	Soil
Type of sample			
Sample Description	-	40g sand, rocks	40g sand, rocks
Asbestos ID in soil	--	No asbestos detected	No asbestos detected
Trace Analysis	--	Respirable fibres not detected	Respirable fibres not detected

Metals in TCLP Our Reference: Your Reference Depth Date Sampled Type of sample	UNITS ----- -----	10350-4 BH1 3-3.45 23/03/07 Soil	10350-10 BH2 3-3.45 23/03/07 Soil	10350-14 BH3 1.1-1.4 23/03/07 Soil	10350-16 BH4 0.5-0.65 23/03/07 Soil	10350-17 BH5 0.1-0.4 24/03/07 Soil
pH of soil for fluid# determ.	pH units	8.20	8.20	9.00	10.50	9.70
pH of soil for fluid # determ. (acid)	pH units	1.50	1.60	1.60	1.70	1.60
Extraction fluid used	-	1	1	1	1	1
pH of final Leachate	pH units	4.90	5.10	5.90	10.40	6.70
Arsenic in TCLP	mg/L	<0.05	<0.05	<0.05	<0.05	<0.05
Cadmium in TCLP	mg/L	<0.01	<0.01	<0.01	<0.01	<0.01
Chromium in TCLP	mg/L	<0.01	<0.01	<0.01	0.05	<0.01
Lead in TCLP	mg/L	<0.03	<0.03	<0.03	<0.03	<0.03
Mercury in TCLP	mg/L	<0.0005	<0.0005	<0.0005	<0.0005	<0.0005
Nickel in TCLP	mg/L	<0.02	0.04	0.03	<0.02	<0.02

Metals in TCLP Our Reference: Your Reference Depth Date Sampled Type of sample	UNITS ----- -----	10350-24 BH7 1.5-1.95 24/03/07 Soil
pH of soil for fluid# determ.	pH units	8.50
pH of soil for fluid # determ. (acid)	pH units	1.50
Extraction fluid used	-	1
pH of final Leachate	pH units	5.00
Arsenic in TCLP	mg/L	<0.05
Cadmium in TCLP	mg/L	<0.01
Chromium in TCLP	mg/L	<0.01
Lead in TCLP	mg/L	0.03
Mercury in TCLP	mg/L	<0.0005
Nickel in TCLP	mg/L	0.03

PAHs in TCLP (USEPA 1311)						
Our Reference:	UNITS	10350-4	10350-10	10350-14	10350-16	10350-17
Your Reference	-----	BH1	BH2	BH3	BH4	BH5
Depth	-----	3-3.45	3-3.45	1.1-1.4	0.5-0.65	0.1-0.4
Date Sampled		23/03/07	23/03/07	23/03/07	23/03/07	24/03/07
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	--	2/04/2007	2/04/2007	2/04/2007	2/04/2007	2/04/2007
Date analysed	--	2/04/2007	2/04/2007	2/04/2007	2/04/2007	2/04/2007
Naphthalene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Acenaphthylene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Acenaphthene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Fluorene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Phenanthrene	mg/L	<0.001	<0.001	0.002	0.002	<0.001
Anthracene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Fluoranthene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Pyrene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Benzo(a)anthracene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Chrysene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Benzo(b,k)fluoranthene	mg/L	<0.002	<0.002	<0.002	<0.002	<0.002
Benzo(a)pyrene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Indeno(1,2,3-c,d)pyrene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Dibenzo(a,h)anthracene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Benzo(g,h,i)perylene	mg/L	<0.001	<0.001	<0.001	<0.001	<0.001
Surrogate p-Terphenyl-d14	%	95	95	89	110	97

PAHs in TCLP (USEPA 1311)	UNITS	10350-24
Our Reference:	-----	BH7
Your Reference	-----	1.5-1.95
Depth		24/03/07
Date Sampled		Soil
Type of sample		
Date extracted	--	2/04/2007
Date analysed	--	2/04/2007
Naphthalene	mg/L	<0.001
Acenaphthylene	mg/L	<0.001
Acenaphthene	mg/L	<0.001
Fluorene	mg/L	<0.001
Phenanthrene	mg/L	<0.001
Anthracene	mg/L	<0.001
Fluoranthene	mg/L	<0.001
Pyrene	mg/L	<0.001
Benzo(a)anthracene	mg/L	<0.001
Chrysene	mg/L	<0.001
Benzo(b,k)fluoranthene	mg/L	<0.002
Benzo(a)pyrene	mg/L	<0.001
Indeno(1,2,3-c,d)pyrene	mg/L	<0.001
Dibenzo(a,h)anthracene	mg/L	<0.001
Benzo(g,h,i)perylene	mg/L	<0.001
Surrogate p-Terphenyl-d14	%	95

sPOCAS Our Reference: Your Reference Depth Date Sampled Type of sample	UNITS ----- -----	10350-28 BH1 2-2.5 23/03/07 Soil	10350-29 BH1 3.5-4 23/03/07 Soil	10350-30 BH5 2.5-2.6 26/03/07 Soil	10350-31 BH5 3.5-4 26/03/07 Soil	10350-32 BH5 4.5-4.7 26/03/07 Soil
pH _{KCl}	pH units	8.1	7.3	8.7	8.1	9.52
TAA pH 6.5	moles H ⁺ /tonne	<5	<5	<5	<5	<5
s-TAA pH 6.5	%w/w S	<0.01	<0.01	<0.01	<0.01	<0.01
pH _{ox}	pH units	7.3	2.4	6.6	2.9	6.5
TPA pH 6.5	moles H ⁺ /tonne	5.0	313	<5.0	25	<5.0
s-TPA pH 6.5	%w/w S	<0.01	0.50	<0.01	0.040	<0.01
TSA pH 6.5	moles H ⁺ /tonne	5.0	313	<5.0	25	<5.0
s-TSA pH 6.5	%w/w S	<0.01	0.50	<0.01	0.040	<0.01
ANCE	% CaCO ₃	0.63	<0.05	0.88	<0.05	0.3
a-ANCE	moles H ⁺ /tonne	125	<5	175	<5	50
s-ANCE	%w/w S	0.20	<0.05	0.28	<0.05	0.08
SKCl	%w/w	0.013	0.058	0.009	0.011	0.070
SP	%w/w	0.052	0.57	0.078	0.077	0.11
SPOS	%w/w	0.039	0.52	0.069	0.066	0.036
a-SPOS	moles H ⁺ /tonne	25	322	43	41	22
CaKCl	%w/w	0.19	0.076	0.23	0.053	0.18
CaP	%w/w	0.31	0.059	0.49	0.049	0.34
CaA	%w/w	0.12	<0.005	0.26	<0.005	0.16
MgKCl	%w/w	0.015	0.017	<0.005	<0.005	<0.005
MgP	%w/w	0.024	0.015	0.026	0.005	0.013
MgA	%w/w	0.009	<0.005	0.022	<0.005	0.011
SRAS	%w/w	<0.005	<0.005	<0.005	<0.005	<0.005
SHCl	%w/w	0.012	0.016	0.009	<0.005	<0.005
SNAS	%w/w	<0.005	<0.005	<0.005	<0.005	<0.005
a-SNAS	moles H ⁺ /tonne	<5	<5	<5	<5	<5
a-SNAS	%w/w S	<0.01	<0.01	<0.01	<0.01	<0.01
a-Net Acidity	moles H ⁺ /tonne	11	315	<10	30	<10
Liming rate	kg CaCO ₃ /tonne	0.83	24	<0.1	2.3	<0.1

Method ID	Methodology Summary
GC.16	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS.
GC.14	Soil samples extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS.
GC.3	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID.
GC.12	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS.
GC-5	Soil samples are extracted with hexane/acetone and waters with dichloromethane and analysed by GC with dual ECD's.
GC-6	Soil samples are extracted with hexane/acetone and waters with dichloromethane and analysed by GC-ECD.
Metals.20	Determination of various metals by ICP-AES.
Metals.21	Determination of Mercury by Cold Vapour AAS.
LAB.8	Moisture content determined by heating at 105 deg C for a minimum of 4 hours.
ASB.1	Qualitative identification of asbestos type fibres in bulk using Polarised Light Microscopy and Dispersion Staining Techniques.
EXTRACT.7	Toxicity Characteristic Leaching Procedure (TCLP).
LAB.64	sPOCAS determined using titrimetric and ICP-AES techniques. Based on Acid Sulfate Soils Laboratory Methods Guidelines, Version 2.1 - June 2004.

QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recover
vTPH & BTEX in Soil						Base II Duplicate II %RPD		
Date extracted	--			[NT]	10350-1	30/03/2007 30/03/2007	[NR]	[NR]
Date analysed	--			[NT]	10350-1	30/03/2007 30/03/2007	[NR]	[NR]
vTPH C6 - C9	mg/kg	25	GC.16	<25	10350-1	<25 <25	LCS	105%
Benzene	mg/kg	1	GC.14	<1.0	10350-1	<1.0 <1.0	LCS	124%
Toluene	mg/kg	1	GC.14	<1.0	10350-1	<1.0 <1.0	LCS	101%
Ethylbenzene	mg/kg	1	GC.14	<1.0	10350-1	<1.0 <1.0	LCS	99%
m + p-Xylene	mg/kg	2	GC.14	<2.0	10350-1	<2.0 <2.0	LCS	100%
o-Xylene	mg/kg	1	GC.14	<1.0	10350-1	<1.0 <1.0	LCS	103%
Surrogate aaa-Trifluorotoluene	%		GC.14	[NT]	10350-1	88 96 RPD: 9	LCS	96%
QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recover
sTPH in Soil (C10-C36)						Base II Duplicate II %RPD		
Date extracted	--			[NT]	10350-1	29/03/2007 29/03/2007	[NR]	[NR]
Date analysed	--			[NT]	10350-1	31/03/2007 31/03/2007	[NR]	[NR]
TPH C10 - C14	mg/kg	50	GC.3	<50	10350-1	<50 <50	LCS	86%
TPH C15 - C28	mg/kg	100	GC.3	<100	10350-1	<100 <100	LCS	72%
TPH C29 - C36	mg/kg	100	GC.3	<100	10350-1	<100 <100	LCS	76%
QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recover
PAHs in Soil						Base II Duplicate II %RPD		
Date extracted	--			[NT]	10350-1	30/03/2007 30/03/2007	[NR]	[NR]
Date analysed	--			[NT]	10350-1	30/03/2007 30/03/2007	[NR]	[NR]
Naphthalene	mg/kg	0.1	GC.12	<0.1	10350-1	<0.1 <0.1	LCS	89%
Acenaphthylene	mg/kg	0.1	GC.12	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Acenaphthene	mg/kg	0.1	GC.12	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Fluorene	mg/kg	0.1	GC.12	<0.1	10350-1	<0.1 <0.1	LCS	85%
Phenanthrene	mg/kg	0.1	GC.12	<0.1	10350-1	0.4 <0.1	LCS	76%
Anthracene	mg/kg	0.1	GC.12	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Fluoranthene	mg/kg	0.1	GC.12	<0.1	10350-1	0.9 0.1 RPD: 160	LCS	93%
Pyrene	mg/kg	0.1	GC.12	<0.1	10350-1	1.0 0.1 RPD: 164	LCS	93%
Benzo(a)anthracene	mg/kg	0.1	GC.12	<0.1	10350-1	0.5 <0.1	[NR]	[NR]
Chrysene	mg/kg	0.1	GC.12	<0.1	10350-1	0.6 <0.1	LCS	95%
Benzo(b,k)fluoranthene	mg/kg	0.2	GC.12	<0.2	10350-1	0.7 <0.2	[NR]	[NR]
Benzo(a)pyrene	mg/kg	0.05	GC.12	<0.05	10350-1	0.5 0.06 RPD: 157	LCS	75%
Indeno(1,2,3-c,d)pyrene	mg/kg	0.1	GC.12	<0.1	10350-1	0.2 <0.1	[NR]	[NR]
Dibenzo(a,h)anthracene	mg/kg	0.1	GC.12	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Benzo(g,h,i)perylene	mg/kg	0.1	GC.12	<0.1	10350-1	0.2 <0.1	[NR]	[NR]
Surrogate p-Terphenyl-d14	%		GC.12	[NT]	10350-1	89 87 RPD: 2	LCS	88%

QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results Base II Duplicate II %RPD	Spike Sm#	Spike % Recovery
Organochlorine Pesticides in soil								
Date extracted	--			[NT]	10350-1	30/03/2007 30/03/2007	[NR]	[NR]
Date analysed	--			[NT]	10350-1	31/03/2007 31/03/2007	[NR]	[NR]
HCB	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
alpha-BHC	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	120%
gamma-BHC	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
beta-BHC	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	131%
Heptachlor	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	120%
delta-BHC	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Aldrin	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	120%
Heptachlor Epoxide	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	125%
gamma-Chlordane	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
alpha-chlordane	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Endosulfan I	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
pp-DDE	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	119%
Dieldrin	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	118%
Endrin	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	66%
pp-DDD	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	127%
Endosulfan II	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
pp-DDT	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Endrin Aldehyde	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Endosulfan Sulphate	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	LCS	109%
Methoxychlor	mg/kg	0.1	GC-5	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Surrogate TCLMX	%		GC-5	[NT]	10350-1	98 96 RPD: 2	LCS	105%

QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recove
PCBs in Soil						Base II Duplicate II %RPD		
Date extracted	--			[NT]	10350-1	30/03/2007 30/03/2007	[NR]	[NR]
Date analysed	--			[NT]	10350-1	31/03/2007 31/03/2007	[NR]	[NR]
Arochlor 1016	mg/kg	0.1	GC-6	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Arochlor 1232	mg/kg	0.1	GC-6	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Arochlor 1242	mg/kg	0.1	GC-6	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Arochlor 1248	mg/kg	0.1	GC-6	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Arochlor 1254	mg/kg	0.1	GC-6	<0.1	10350-1	<0.1 <0.1	LCS	107%
Arochlor 1260	mg/kg	0.1	GC-6	<0.1	10350-1	<0.1 <0.1	[NR]	[NR]
Surrogate TCLMX	%		GC-6	[NT]	10350-1	98 96 RPD: 2	LCS	82%
QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recove
Acid Extractable metals in soil						Base II Duplicate II %RPD		
Arsenic	mg/kg	4	Metals.20	<4.0	10350-1	<4.0 <4.0	[NR]	[NR]
Cadmium	mg/kg	1	Metals.20	<1.0	10350-1	<1.0 <1.0	[NR]	[NR]
Chromium	mg/kg	1	Metals.20	<1.0	10350-1	10 11 RPD: 10	[NR]	[NR]
Copper	mg/kg	1	Metals.20	<1.0	10350-1	30 30 RPD: 0	[NR]	[NR]
Lead	mg/kg	1	Metals.20	<1.0	10350-1	14 16 RPD: 13	[NR]	[NR]
Mercury	mg/kg	0.1	Metals.21	<0.10	10350-1	0.20 <0.10	[NR]	[NR]
Nickel	mg/kg	1	Metals.20	<1.0	10350-1	33 34 RPD: 3	[NR]	[NR]
Zinc	mg/kg	1	Metals.20	<1.0	10350-1	39 44 RPD: 12	[NR]	[NR]
QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results		
Moisture						Base II Duplicate II %RPD		
Moisture	%	0.1	LAB.8	<0.10	10350-1	8.4 8.4 RPD: 0		
QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recove
Metals in TCLP						Base II Duplicate II %RPD		
Arsenic in TCLP	mg/L	0.05	Metals.20	<0.05	[NT]	[NT]	LCS	111%
Cadmium in TCLP	mg/L	0.01	Metals.20	<0.01	[NT]	[NT]	LCS	102%
Chromium in TCLP	mg/L	0.01	Metals.20	<0.01	[NT]	[NT]	LCS	104%
Lead in TCLP	mg/L	0.03	Metals.20	<0.03	[NT]	[NT]	LCS	101%
Mercury in TCLP	mg/L	0.0005	Metals.21	<0.0005	[NT]	[NT]	LCS	105%
Nickel in TCLP	mg/L	0.02	Metals.20	<0.02	[NT]	[NT]	LCS	103%
QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recove
PAHs in TCLP (USEPA 1311)						Base II Duplicate II %RPD		
Date extracted	--			[NT]	[NT]	[NT]	[NR]	[NR]
Date analysed	--			[NT]	[NT]	[NT]	[NR]	[NR]
Naphthalene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	LCS	104%
Acenaphthylene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	[NR]	[NR]
Acenaphthene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	[NR]	[NR]
Fluorene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	LCS	95%

QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recove
PAHs in TCLP (USEPA 1311)						Base II Duplicate II %RPD		
Phenanthrene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	LCS	87%
Anthracene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	[NR]	[NR]
Fluoranthene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	LCS	102%
Pyrene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	LCS	98%
Benzo(a)anthracene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	[NR]	[NR]
Chrysene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	LCS	98%
Benzo(b,k)fluoranthene	mg/L	0.002	GC.12	<0.002	[NT]	[NT]	[NR]	[NR]
Benzo(a)pyrene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	LCS	95%
Indeno(1,2,3-c,d)pyrene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	[NR]	[NR]
Dibenzo(a,h)anthracene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	[NR]	[NR]
Benzo(g,h,i)perylene	mg/L	0.001	GC.12	<0.001	[NT]	[NT]	[NR]	[NR]
Surrogate p-Terphenyl-d14	%		GC.12	[NT]	[NT]	[NT]	LCS	115%
QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recove
sPOCAS						Base II Duplicate II %RPD		
pH _{KCl}	pH units		LAB.64	[NT]	[NT]	[NT]	LCS	99%
TAA pH 6.5	moles H ⁺ /tonne	5	LAB.64	<5	[NT]	[NT]	LCS	119%
s-TAA pH 6.5	%w/w S	0.01	LAB.64	<0.01	[NT]	[NT]	[NR]	[NR]
pH _{ox}	pH units		LAB.64	[NT]	[NT]	[NT]	LCS	98%
TPA pH 6.5	moles H ⁺ /tonne	5	LAB.64	<5.0	[NT]	[NT]	LCS	106%
s-TPA pH 6.5	%w/w S	0.01	LAB.64	<0.01	[NT]	[NT]	[NR]	[NR]
TSA pH 6.5	moles H ⁺ /tonne	5	LAB.64	<5.0	[NT]	[NT]	[NR]	[NR]
s-TSA pH 6.5	%w/w S	0.01	LAB.64	<0.01	[NT]	[NT]	[NR]	[NR]
ANCE	% CaCO ₃	0.05	LAB.64	<0.05	[NT]	[NT]	[NR]	[NR]
a-ANCE	moles H ⁺ /tonne	5	LAB.64	<5	[NT]	[NT]	[NR]	[NR]
s-ANCE	%w/w S	0.05	LAB.64	<0.05	[NT]	[NT]	[NR]	[NR]
SKCl	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	LCS	141%
SP	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	LCS	109%
SPOS	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	[NR]	[NR]
a-SPOS	moles H ⁺ /tonne	5	LAB.64	<5.0	[NT]	[NT]	[NR]	[NR]

QUALITY CONTROL	UNITS	PQL	METHOD	Blank	Duplicate Sm#	Duplicate results	Spike Sm#	Spike % Recovery
sPOCAS						Base Duplicate %RPD		
CaKCl	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	LCS	77%
CaP	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	LCS	77%
CaA	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	[NR]	[NR]
MgKCl	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	LCS	79%
MgP	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	LCS	122%
MgA	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	[NR]	[NR]
SRAS	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	[NR]	[NR]
SHCl	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	LCS	97%
SNAS	%w/w	0.005	LAB.64	<0.005	[NT]	[NT]	[NR]	[NR]
a-SNAS	moles H ⁺ /tonne	5	LAB.64	<5	[NT]	[NT]	[NR]	[NR]
a-SNAS	%w/w S	0.01	LAB.64	<0.01	[NT]	[NT]	[NR]	[NR]
a-Net Acidity	moles H ⁺ /tonne	10	LAB.64	<10	[NT]	[NT]	[NR]	[NR]
Liming rate	kg CaCO ₃ /tonne	0.1	LAB.64	<0.1	[NT]	[NT]	[NR]	[NR]
QUALITY CONTROL	UNITS	Dup. Sm#		Duplicate		Spike Sm#	Spike % Recovery	
vTPH & BTEX in Soil				Base + Duplicate + %RPD				
Date extracted	--	10350-22		30/03/2007 30/03/2007		[NR]	[NR]	
Date analysed	--	10350-22		30/03/2007 30/03/2007		[NR]	[NR]	
vTPH C ₆ - C ₉	mg/kg	10350-22		<25 <25		10350-4	95%	
Benzene	mg/kg	10350-22		<1.0 <1.0		10350-4	97%	
Toluene	mg/kg	10350-22		<1.0 <1.0		10350-4	95%	
Ethylbenzene	mg/kg	10350-22		<1.0 <1.0		10350-4	93%	
m + p-Xylene	mg/kg	10350-22		<2.0 <2.0		10350-4	93%	
o-Xylene	mg/kg	10350-22		<1.0 <1.0		10350-4	96%	
Surrogate aaa-Trifluorotoluene	%	10350-22		78 94 RPD: 19		10350-4	95%	

QUALITY CONTROL sTPH in Soil (C10-C36)	UNITS	Dup. Sm#	Duplicate Base + Duplicate + %RPD	Spike Sm#	Spike % Recovery
Date extracted	--	10350-22	29/03/2007 29/03/2007	[NR]	[NR]
Date analysed	--	10350-22	31/03/2007 31/03/2007	[NR]	[NR]
TPH C10 - C14	mg/kg	10350-22	<50 <50	10350-4	87%
TPH C15 - C28	mg/kg	10350-22	<100 <100	10350-4	71%
TPH C29 - C36	mg/kg	10350-22	<100 <100	10350-4	73%
QUALITY CONTROL PAHs in Soil	UNITS	Dup. Sm#	Duplicate Base + Duplicate + %RPD	Spike Sm#	Spike % Recovery
Date extracted	--	10350-22	30/03/2007 30/03/2007	[NR]	[NR]
Date analysed	--	10350-22	30/03/2007 30/03/2007	[NR]	[NR]
Naphthalene	mg/kg	10350-22	<0.1 <0.1	10350-4	78%
Acenaphthylene	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Acenaphthene	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Fluorene	mg/kg	10350-22	<0.1 <0.1	10350-4	80%
Phenanthrene	mg/kg	10350-22	<0.1 <0.1	10350-4	60%
Anthracene	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Fluoranthene	mg/kg	10350-22	<0.1 <0.1	10350-4	87%
Pyrene	mg/kg	10350-22	<0.1 <0.1	10350-4	84%
Benzo(a)anthracene	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Chrysene	mg/kg	10350-22	<0.1 <0.1	10350-4	82%
Benzo(b,k)fluoranthene	mg/kg	10350-22	<0.2 <0.2	[NR]	[NR]
Benzo(a)pyrene	mg/kg	10350-22	<0.05 <0.05	10350-4	76%
Indeno(1,2,3-c,d)pyrene	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Dibenzo(a,h)anthracene	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Benzo(g,h,i)perylene	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Surrogate p-Terphenyl-d14	%	10350-22	88 88 RPD: 0	10350-4	88%

QUALITY CONTROL Organochlorine Pesticides in soil	UNITS	Dup. Sm#	Duplicate Base + Duplicate + %RPD	Spike Sm#	Spike % Recovery
Date extracted	--	10350-22	30/03/2007 30/03/2007	[NR]	[NR]
Date analysed	--	10350-22	31/03/2007 31/03/2007	[NR]	[NR]
HCB	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
alpha-BHC	mg/kg	10350-22	<0.1 <0.1	10350-4	111%
gamma-BHC	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
beta-BHC	mg/kg	10350-22	<0.1 <0.1	10350-4	124%
Heptachlor	mg/kg	10350-22	<0.1 <0.1	10350-4	112%
delta-BHC	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Aldrin	mg/kg	10350-22	<0.1 <0.1	10350-4	115%
Heptachlor Epoxide	mg/kg	10350-22	<0.1 <0.1	10350-4	117%
gamma-Chlordane	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
alpha-chlordane	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Endosulfan I	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
pp-DDE	mg/kg	10350-22	<0.1 <0.1	10350-4	114%
Dieldrin	mg/kg	10350-22	<0.1 <0.1	10350-4	111%
Endrin	mg/kg	10350-22	<0.1 <0.1	10350-4	112%
pp-DDD	mg/kg	10350-22	<0.1 <0.1	10350-4	122%
Endosulfan II	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
pp-DDT	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Endrin Aldehyde	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Endosulfan Sulphate	mg/kg	10350-22	<0.1 <0.1	10350-4	103%
Methoxychlor	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Surrogate TCLMX	%	10350-22	100 100 RPD: 0	10350-4	98%

QUALITY CONTROL PCBs in Soil	UNITS	Dup. Sm#	Duplicate Base + Duplicate + %RPD	Spike Sm#	Spike % Recovery
Date extracted	--	10350-22	30/03/2007 30/03/2007	[NR]	[NR]
Date analysed	--	10350-22	31/03/2007 31/03/2007	[NR]	[NR]
Arochlor 1016	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Arochlor 1232	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Arochlor 1242	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Arochlor 1248	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Arochlor 1254	mg/kg	10350-22	<0.1 <0.1	10350-4	92%
Arochlor 1260	mg/kg	10350-22	<0.1 <0.1	[NR]	[NR]
Surrogate TCLMX	%	10350-22	100 100 RPD: 0	10350-4	76%
QUALITY CONTROL Acid Extractable metals in soil	UNITS	Dup. Sm#	Duplicate Base + Duplicate + %RPD	Spike Sm#	Spike % Recovery
Arsenic	mg/kg	10350-22	<4.0 <4.0	[NR]	[NR]
Cadmium	mg/kg	10350-22	<1.0 <1.0	[NR]	[NR]
Chromium	mg/kg	10350-22	15 17 RPD: 12	[NR]	[NR]
Copper	mg/kg	10350-22	48 55 RPD: 14	[NR]	[NR]
Lead	mg/kg	10350-22	10 9.5 RPD: 5	[NR]	[NR]
Mercury	mg/kg	10350-22	<0.10 <0.10	[NR]	[NR]
Nickel	mg/kg	10350-22	86 90 RPD: 5	[NR]	[NR]
Zinc	mg/kg	10350-22	45 45 RPD: 0	[NR]	[NR]
QUALITY CONTROL Moisture	UNITS	Dup. Sm#	Duplicate Base + Duplicate + %RPD		
Moisture	%	10350-22	7.9 7.9 RPD: 0		

Report Comments:

PAH's in soil: #1 duplicate RPD is not withing accepted criteria, however the result will be accepted as sample is not homogenous.

Trace Elements: high %RPD duplicate results for Mercury sample 1. This is due to the non homogeneous nature of the sample for this particular element.

Asbestos analysed by: Tania Notaras

INS: Insufficient sample for this test

NT: Not tested

PQL: Practical Quantitation Limit

RPD: Relative Percent Difference

NA: Test not required

LCS: Laboratory Control Sample

NR: Not requested

<: Less than

>: Greater than

Quality Control Definitions

Blank: This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.

Duplicate: This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.

Matrix Spike: A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.

LCS (Laboratory Control Sample): This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.

Surrogate Spike: Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.

Laboratory Acceptance Criteria:

Duplicates: <5xPQL - any RPD is acceptable;

>5xPQL - 0-50% RPD is acceptable.

Matrix Spikes and LCS: Generally 70-130% for inorganics/metals; 60-140% for organics and 10-140% for SVOC and speciated phenols is acceptable.

Surrogates: Generally 60-140% is acceptable.



Envirolab Services Pty Ltd

ABN 37 112 535 645

54 Frenchs Rd Willoughby NSW 2068

ph 02 9958 5801 fax 02 9958 5803

email: tnotaras@envirolabservices.com.au

SAMPLE RECEIPT ADVICE

Client:

Environmental Investigation Services

PO Box 976

North Ryde BC NSW 1670

ph: 02 9888 5000

Fax: 02 9888 5001

Attention: Todd Hore

Sample log in details:

Your reference:

E21073F, Darling Harbour

Envirolab Reference:

10350

Date received:

27/03/07

Date results expected to be reported:

3/04/07

Samples received in appropriate condition for analysis:

YES

Turnaround time requested:

Standard

Temperature on receipt

Cool

Cooling Method:

Ice Pack

Completed documentation received:

YES

Comments:

Samples will be held for 1 month for water samples and 2 months for soil samples from date of receipt of samples.

Contact details:

Please direct any queries to Aileen Hie or Jacinta Hurst

ph: 02 9958 5801 fax: 02 9958 5803

email: ahie@envirolabservices.com.au or jhurst@envirolabservices.com.au

SAMPLE AND CHAIN OF CUSTODY FORM

TO: Envirolab Services Pty Ltd 54 Frenchs Road Willoughby NSW 2068 Phone: (02) 9958 5801 Fax: (02) 9958 5803 Attention: Aileen				EIS Job Number: E21073F Date Results Required: standard turnaround				FROM: Environmental Investigation Services Rear 115 Wicks Road Macquarie Park NSW 2113 Phone: (02) 9888 5000 Fax: (02) 9888 5004 Contact: Todd Hore									
Project: Proposed Commercial Development Location: Darling Harbour Sampler: TH				Tests Required				Sample Preservation: In esky on ice									
Date Sampled	Lab Ref:	Borehole/ Sample Number	Depth (m)	Sample Container	PID	Sample Description	Combo #5	Combo #5a	Combo #12	Combo #12a	Asbestos	Heavy metals	TPH/ BTEX	PAH	TCLP Prep +		
1 23/3/07		BH1	0.1-0.3	Glass jar + Asb Bag	0	Soil		X									
2 23/3/07		BH1	0.5-0.95	Glass jar + Asb Bag	0	Soil											
3 23/3/07		BH1	1.5-1.95	Glass jar + Asb Bag	0	Soil											
4 23/3/07		BH1	3-3.45	Glass jar + Asb Bag	0	Soil				X							
5 23/3/07		BH1	4-4.3	Glass jar	0	Soil											
6 23/3/07		BH1	4.5-4.67	Glass jar + Asb Bag	0	Rock											
7 23/3/07		BH2	0.1-0.4	Glass jar + Asb Bag	0	Soil											
8 23/3/07		BH2	0.5-0.95	Glass jar + Asb Bag	0	Soil		X									
9 23/3/07		BH2	1.5-1.95	Glass jar + Asb Bag	0	Soil											
10 23/3/07		BH2	3-3.45	Glass jar + Asb Bag	0	Soil				X							
11 23/3/07		BH2	4.4-4.7	Glass jar + Asb Bag	0	Rock											
12 23/3/07		BH3	0.1-0.4	Glass jar + Asb Bag	0	Soil		X									
13 23/3/07		BH3	0.5-0.64	Glass jar + Asb Bag	0	Soil											
14 23/3/07		BH3	1.1-1.4	Glass jar + Asb Bag	0	Soil				X							
15 23/3/07		BH4	0.1-0.5	Glass jar + Asb Bag	0	Soil		X									
16 23/3/07		BH4	0.5-0.65	Glass jar + Asb Bag	0	Soil				X							
17 24/3/07		BH5	0.1-0.4	Glass jar + Asb Bag	0	Soil				X							
18 24/3/07		BH5	0.5-0.95	Glass jar + Asb Bag	0	Soil											
19 24/3/07		BH5	1.6-1.85	Glass jar + Asb Bag	0	Soil		X									
20 24/3/07		BH5	3-3.45	Glass jar + Asb Bag	0	Soil											
21 24/3/07		BH5	4.7-4.95	Glass jar + Asb Bag	0	Soil											
22 24/3/07		BH7	0.1-0.4	Glass jar + Asb Bag	0	Soil		X									
23 24/3/07		BH7	0.5-0.95	Glass jar + Asb Bag	0	Soil											
24 24/3/07		BH7	1.5-1.95	Glass jar + Asb Bag	0	Soil				X							
25 23/3/07		Dup 3		Glass jar	0	Soil	X										

Envirolab Services
 54 Frenchs Rd
 Willoughby NSW 2068
 Ph: 9958 5801
 Job No: 10350
 Date received: 27/3/07
 Received by: [Signature]
 Temp: Cool/Ambient
 Security: Intact/Broken/None

Remarks (comments/detection limits required):

Relinquished By: [Signature]	Date: 27/3/07 Time: 12pm	Received By: [Signature]	Relinquished By:	Date:	Received By:
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SAMPLE AND CHAIN OF CUSTODY FORM

[illegible]

APPENDIX C



espreon
tomorrow. today™

Date 11.4.2007

Topic Historical

People

Search Report

Re: Darling Walk, Darling Harbour

1. 318/871455 Title

318/871455 Historical

2. Prior title

303/787105 Historical

3. Prior title - 204/771840 Historical

AP 871455, AP 787105, AP 771840, AP 738093

(Copies attached).

Note: There are numerous prior titles for
Historical on 204/771840, Please advise if
further searches are required.

[Signature]



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Information provided through Tri-Search an approved LPI/NSW Information Broker

LAND AND PROPERTY INFORMATION NEW SOUTH WALES - TITLE SEARCH

FOLIO: 318/871455

SEARCH DATE	TIME	EDITION NO	DATE
11/4/2007	3:41 PM	6	26/5/2006

LAND

LOT 318 IN DEPOSITED PLAN 871455
AT DARLING HARBOUR
LOCAL GOVERNMENT AREA SYDNEY
PARISH OF ST ANDREW COUNTY OF CUMBERLAND
TITLE DIAGRAM DP871455

FIRST SCHEDULE

SYDNEY HARBOUR FORESHORE AUTHORITY (CN 8554459)

SECOND SCHEDULE (4 NOTIFICATIONS)

- 1 RESERVATIONS AND CONDITIONS IN THE CROWN GRANT(S)
- 2 DP869004 EASEMENT FOR STORMWATER DRAINAGE 4.5 WIDE AFFECTING THE PARTS SHOWN SO BURDENED IN DP869004
- 3 DP869004 EASEMENT FOR STORMWATER DRAINAGE 11.4 WIDE AFFECTING THE PART(S) SHOWN SO BURDENED IN DP869004
- 4 5060885 LEASE TO JACFUN PTY LIMITED EXPIRES: 21/3/2038.
OPTION OF RENEWAL: TWO PERIODS OF 26 YEARS EACH.
6245470 LEASE OF LEASE 5060885 TO MCDONALD'S PROPERTIES (AUSTRALIA) PTY LIMITED OF SHOP 9, THE CORRAL AREA AND THE REMOTE BOOTH AS SHOWN IN PLAN WITH 6245470. EXPIRES: 19/9/2006. OPTION OF RENEWAL: 7 YEARS WITH A FURTHER PERIOD OF 7 YEARS.
7117607 LEASE OF LEASE 5060885 TO JIREH INTERNATIONAL PTY LIMITED OF SHOPS 8 & 10, "DARLING WALK" DARLING HARBOUR. EXPIRES: 20/8/2010. OPTION OF RENEWAL: 5 YEARS.
7155336 VARIATION OF LEASE 5060885
* 9520184 CAVEAT AFFECTING LEASE 5060885 CAVEAT BY MCDONALDS PROPERTIES (AUSTRALIA) PTY LIMITED OF PART BEING SHOP 9, THE CORRAL AREA & THE REMOTE BOOTH SHOWN HATCHED IN PLAN WITH 6245470

NOTATIONS

UNREGISTERED DEALINGS: NIL

*** END OF SEARCH ***

jg

PRINTED ON 11/4/2007



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LAND AND PROPERTY INFORMATION NEW SOUTH WALES - HISTORICAL SEARCH

SEARCH DATE

11/4/2007 3:42PM

FOLIO: 318/871455

First Title(s): OLD SYSTEM

Prior Title(s): 303/787105

Recorded	Number	Type of Instrument	C.T. Issue
18/9/1997	DP871455	DEPOSITED PLAN	FOLIO CREATED EDITION 1
8/7/1998	5061036	WITHDRAWAL OF CAVEAT	
8/7/1998	5061037	WITHDRAWAL OF CAVEAT	
8/7/1998	5060885	LEASE	
8/7/1998	5060886	MORTGAGE OF LEASE	
8/7/1998	5061038	CAVEAT	
8/7/1998	5060887	SUB-LEASE	EDITION 2
7/9/1998	5248031	SUB-LEASE	
23/9/1998	5284347	SUB-LEASE	
27/5/1999	5856672	SUB-LEASE	
16/7/1999	5989931	DISCHARGE OF MORTGAGE	
7/10/1999	6223893	SUB-LEASE	
13/10/1999	6251866	SUB-LEASE	
26/10/1999	6276320	SUB-LEASE	
15/11/1999	6245470	SUB-LEASE	
13/12/1999	6415553	SUB-LEASE	
13/12/1999	6415554	SUB-LEASE	
13/12/1999	6415555	SUB-LEASE	
13/12/1999	6415556	SUB-LEASE	
24/12/1999	6450271	SUB-LEASE	
20/1/2000	6500896	SUB-LEASE	
21/1/2000	6504174	SUB-LEASE	
18/10/2000	7117607	SUB-LEASE	
18/10/2000	7120832	SUB-LEASE	
18/10/2000	7155336	VARIATION OF LEASE	EDITION 3

END OF PAGE 1 - CONTINUED OVER

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PRINTED ON 11/4/2007

LAND AND PROPERTY INFORMATION NEW SOUTH WALES - HISTORICAL SEARCH
-----SEARCH DATE

11/4/2007 3:42PM

FOLIO: 318/871455

PAGE 2

Recorded -----	Number -----	Type of Instrument -----	C.T. Issue -----
1/11/2000	7191432	SUB-LEASE	
9/11/2000	7198200	SUB-LEASE	
7/5/2002	8554459	CHANGE OF NAME	
7/5/2002	8571166	DEPARTMENTAL DEALING	EDITION 4
27/6/2002	8720584	CHANGE OF NAME	
27/6/2002	8720585	VARIATION OF LEASE	
3/3/2003	9417374	DETERMINATION OF LEASE	
16/5/2003	9410995	TRANSFER OF LEASE	
5/6/2003	9520184	CAVEAT	
28/6/2004	AA270452	REJECTED - DETERMINATION OF LEASE	
28/6/2004	AA270453	REJECTED - DETERMINATION OF LEASE	
28/6/2004	AA542043	REJECTED - REQUEST	
29/6/2004	AA270451	WITHDRAWAL OF CAVEAT	
30/8/2005	AB721568	REQUEST	
30/8/2005	AB721569	REQUEST	
30/8/2005	AB721570	REQUEST	EDITION 5
24/5/2006	AC294450	WITHDRAWN - LEASE	
24/5/2006	AC294451	WITHDRAWN - LEASE	
26/5/2006	AC294449	REQUEST	EDITION 6

*** END OF SEARCH ***

jg

PRINTED ON 11/4/2007



Lawpoint Galloways hereby certifies that the information contained in this document has been provided electronically by the Registrar General.

Information provided through Tri-Search an approved LPI/NSW Information Broker

LAND AND PROPERTY INFORMATION NEW SOUTH WALES - HISTORICAL SEARCH

SEARCH DATE

11/4/2007 3:42PM

FOLIO: 303/787105

First Title(s): OLD SYSTEM VOL 4577 FOL 244
VOL 489 FOL 80
Prior Title(s): 204/771840

Recorded	Number	Type of Instrument	C.T. Issue
25/10/1989	DP787105	DEPOSITED PLAN	FOLIO CREATED EDITION 1
27/11/1995		AMENDMENT: TITLE DIAGRAM	
19/12/1995	0713447	CAVEAT	
19/12/1995	0713448	CAVEAT	
7/11/1996	2571756	WITHDRAWAL OF CAVEAT	
7/11/1996	2571757	WITHDRAWAL OF CAVEAT	
7/11/1996	2571758	CAVEAT	
7/11/1996	2571759	CAVEAT	
29/5/1997	3104624	CAVEAT	
5/6/1997	DP869004	DEPOSITED PLAN	EDITION 2
18/9/1997	DP871455	DEPOSITED PLAN	FOLIO CANCELLED

*** END OF SEARCH ***

jg

PRINTED ON 11/4/2007



Lawpoint Galloways hereby certifies that the information contained in this document has been provided electronically by the Registrar General.

Information provided through Tri-Search an approved LPI/NSW Information Broker

LAND AND PROPERTY INFORMATION NEW SOUTH WALES - HISTORICAL SEARCH

SEARCH DATE

11/4/2007 3:43PM

FOLIO: 204/771840

First Title(s): OLD SYSTEM VOL 20 FOL 88
VOL 99 FOL 142 VOL 489 FOL 79
VOL 489 FOL 80 VOL 4577 FOL 244
Prior Title(s): VOL 662 FOL 170 VOL 1307 FOLS 63-64
VOL 1374 FOLS 167-168 VOL 1537 FOL 191
VOL 1579 FOL 171 VOL 2223 FOL 102
VOL 2374 FOL 212 VOL 2435 FOLS 215-218
VOL 2856 FOL 221 VOL 3163 FOL 212
VOL 3552 FOL 101 VOL 4522 FOL 91
VOL 4602 FOL 220 VOL 5011 FOL 170
VOL 5770 FOL 135 VOL 5809 FOL 238
VOL 11597 FOL 1 VOL 13061 FOL 226
VOL 14730 FOLS 65-66 VOL 15218 FOL 191
VOL 15231 FOL 105 VOL 15457 FOL 192
PA59783

Recorded	Number	Type of Instrument	C.T. Issue
22/2/1988	PA59783	PRIMARY APPLICATION	FOLIO CREATED EDITION 1
24/10/1989	DP787105	DEPOSITED PLAN	FOLIO CANCELLED

PRIOR TITLES(S) AS AMENDED:

VOL 662 FOL 170, VOL 1307 FOLS 63-64,
VOL 1374 FOLS 167-168, VOL 1537 FOL 191,
VOL 1579 FOL 171, VOL 2223 FOL 102,
VOL 2374 FOL 212, VOL 2435 FOLS 215-218,
VOL 2856 FOL 221, VOL 3163 FOL 212,
VOL 3552 FOL 101, VOL 4552 FOL 91, VOL 4602 FOL 220,
VOL 5011 FOL 170, VOL 5245 FOL 7, VOL 5770 FOL 135,
VOL 5809 FOL 238, VOL 11597 FOL 1, VOL 12946 FOL 247,
VOL 13061 FOL 226, VOL 14730 FOLS 65-66,
VOL 15218 FOL 191, VOL 15231 FOL 105,
VOL 15457 FOL 192, PA59783.

15/3/1996	2018316	DEPARTMENTAL DEALING	FOLIO CANCELLED
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*** END OF SEARCH ***

jg

PRINTED ON 11/4/2007

PLAN FORM 2

SIGNATURE AND SEALS ONLY

Plan Drawing only to appear in this space

OFFICE USE ONLY

DP 787105

Registered 28-4-1994

C.A. TORRENS

SYDNEY SHEET 101

PLAN OF SUBDIVISION OF LOT 204 DP 771860 & LOT 1 DP 714328 & LAND TO BE ACQUIRED.

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PLAN OF SUBDIVISION OF LOT 204 DP 771860 & LOT 1 DP 714328 & LAND TO BE ACQUIRED.

SIGNED BY THE ACTING GENERAL MANAGER UNDER DELEGATED AUTHORITY FOR THE DARLING HARBOUR AUTHORITY.

S. Campbell
for J. O. STANLEY

Crown Lands Office Approval

PLAN APPROVED: Authorised Officer

Land District: ...

Page No: ...

Plan Book: ...

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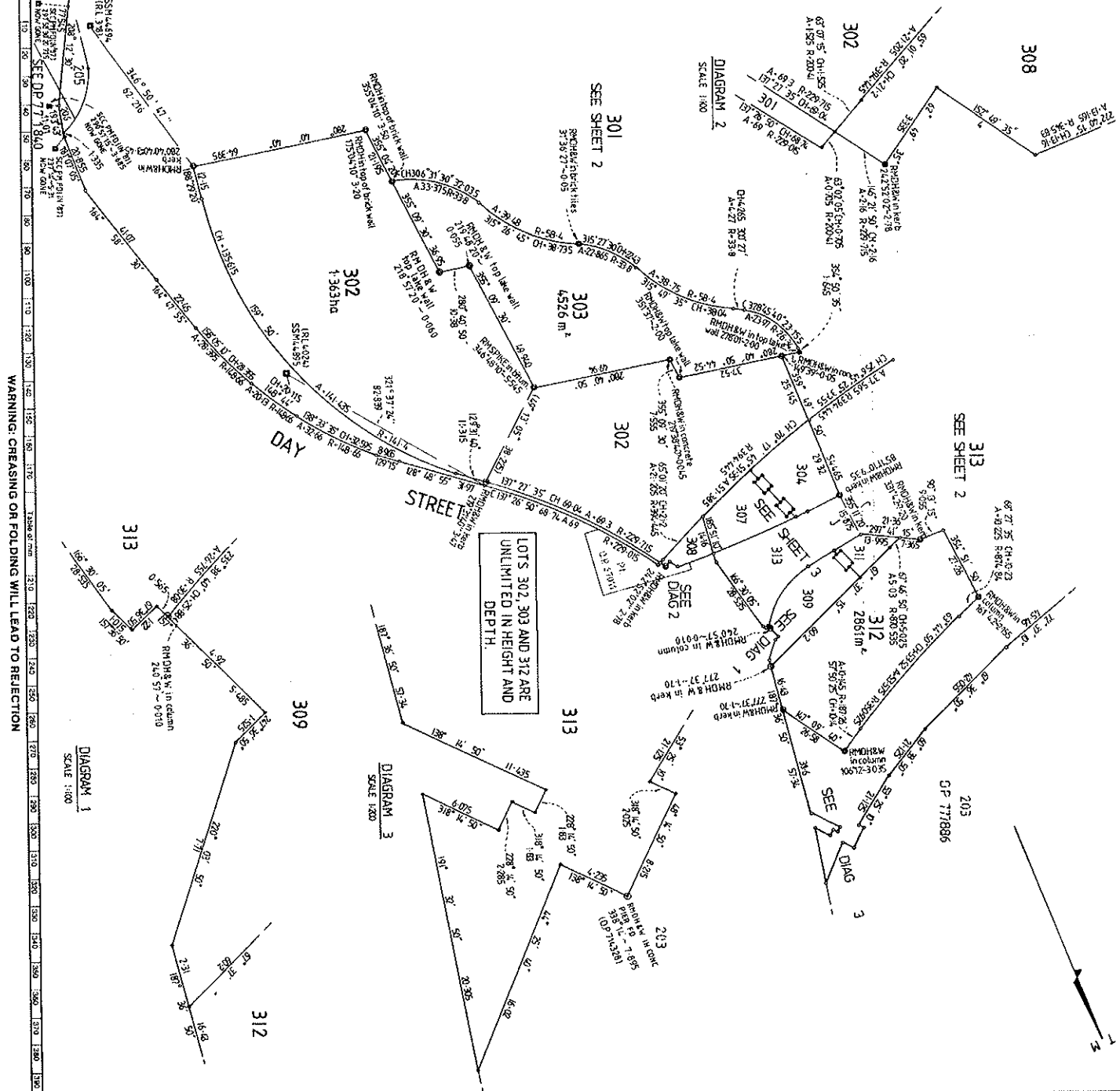
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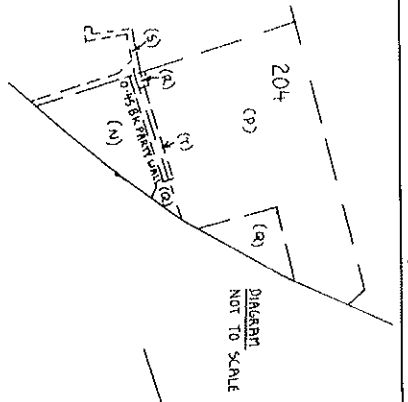
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WARNING: CREASING OR FOLDING WILL LEAD TO REJECTION

SUBDIVISION REFERENCE SD 0812

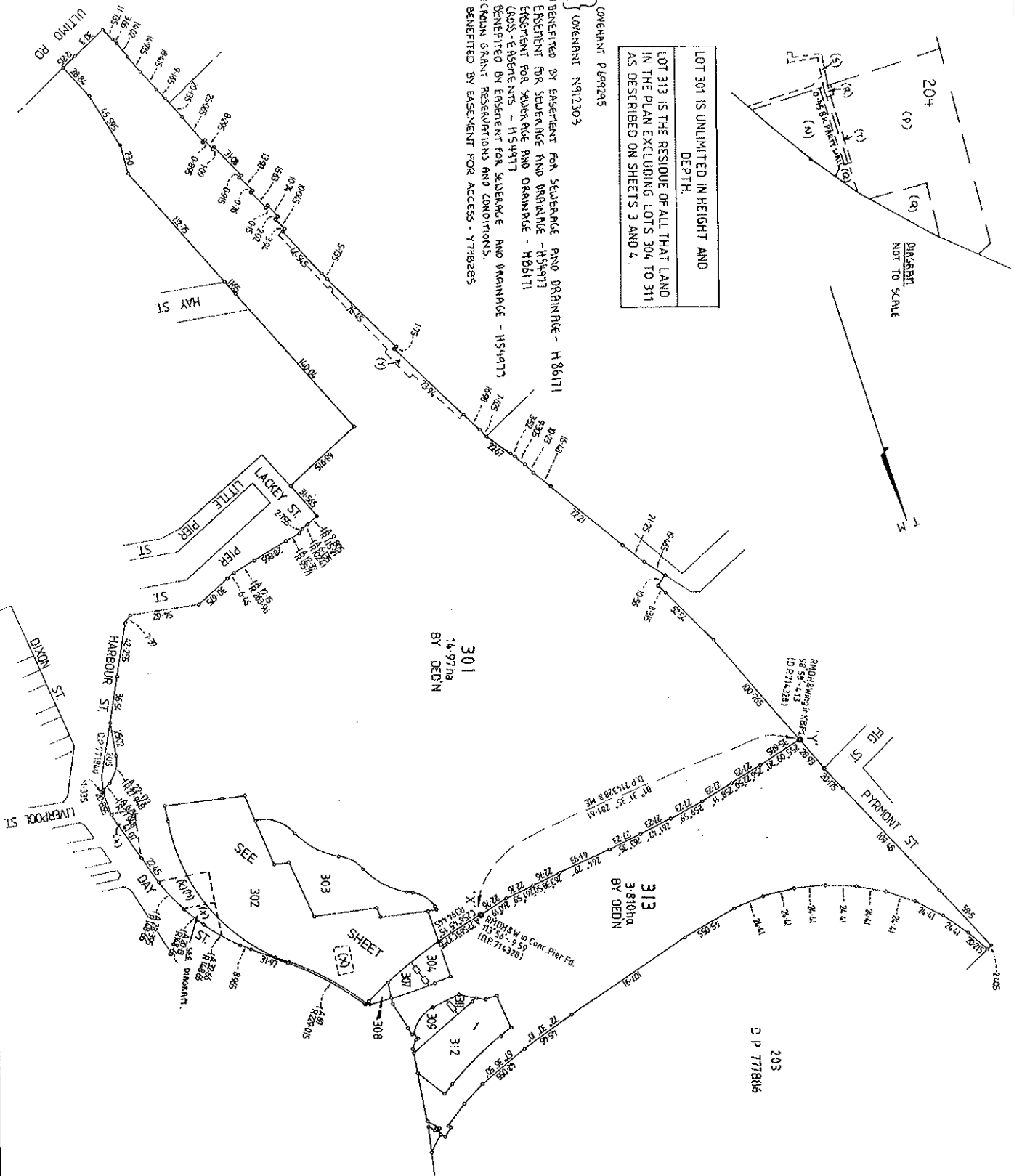
PLAN OF SUBDIVISION OF LOT 204 DP 771860 & LOT 1 DP 714328 & LAND TO BE ACQUIRED.



LOT 301 IS UNLIMITED IN HEIGHT AND DEPTH.

(M) COVENANT P 649295
 (N) COVENANT N 912303
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- (Z) COVENANT N 912303



Plan Drawing only to appear in this space

Surveyor's Reference: SD 0812

DP 787105

Registered: (CAU) 28-9-1997

This is a true and correct copy of the original plan as shown on 23rd JANUARY 1997

Surveyor's Reference: SD 0812

This is a true and correct copy of the original plan as shown on 23rd JANUARY 1997

This is a true and correct copy of the original plan as shown on 23rd JANUARY 1997

For use where stated is a modification in any plan on Form 2

LOTS 304 TO 311 ARE LIMITED IN HEIGHT BY THE INCLINED PLANES BEING THE UNDERSIDE OF LOT 313 AS IDENTIFIED IN THE CROSS SECTIONS ON SHEETS 3 AND 4. AND ARE UNLIMITED IN DEPTH.

DIAGRAM 4
SCALE 1:250

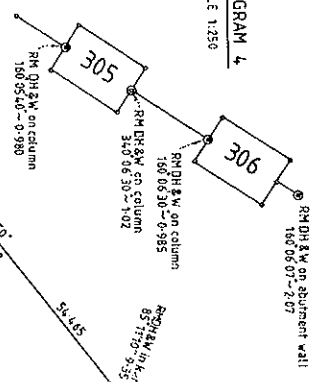
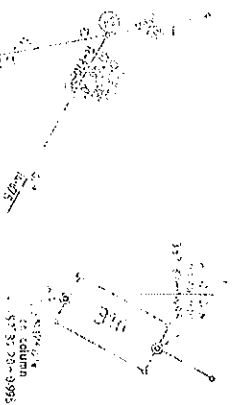


DIAGRAM 5
SCALE 1:250



NOTE: LOT 313 IS ABOVE LOTS 304 TO 311.

302
SEE SHEET 1

312
SEE SHEET 1

SEE
DIAG
1

Plan Drawing only to appear in this space

DP 787105

Registered: CMA 28-4-1988

Trill & Sons 3 of 4 Plan 28-4-1988

dated 2312 JANUARY 1985

U.S. Surveyors

Surveyor registered under Surveyors Act 1980

This is a plan of the land

shown covered by my Certificate No

of

Council Order

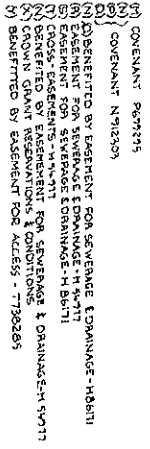
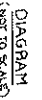
For use under a license is invalid in any other court than

Survey 2

PLAN AMENDED AT
SURVEYORS REQUEST
12.05.97

Reduction Ratio: 1:250

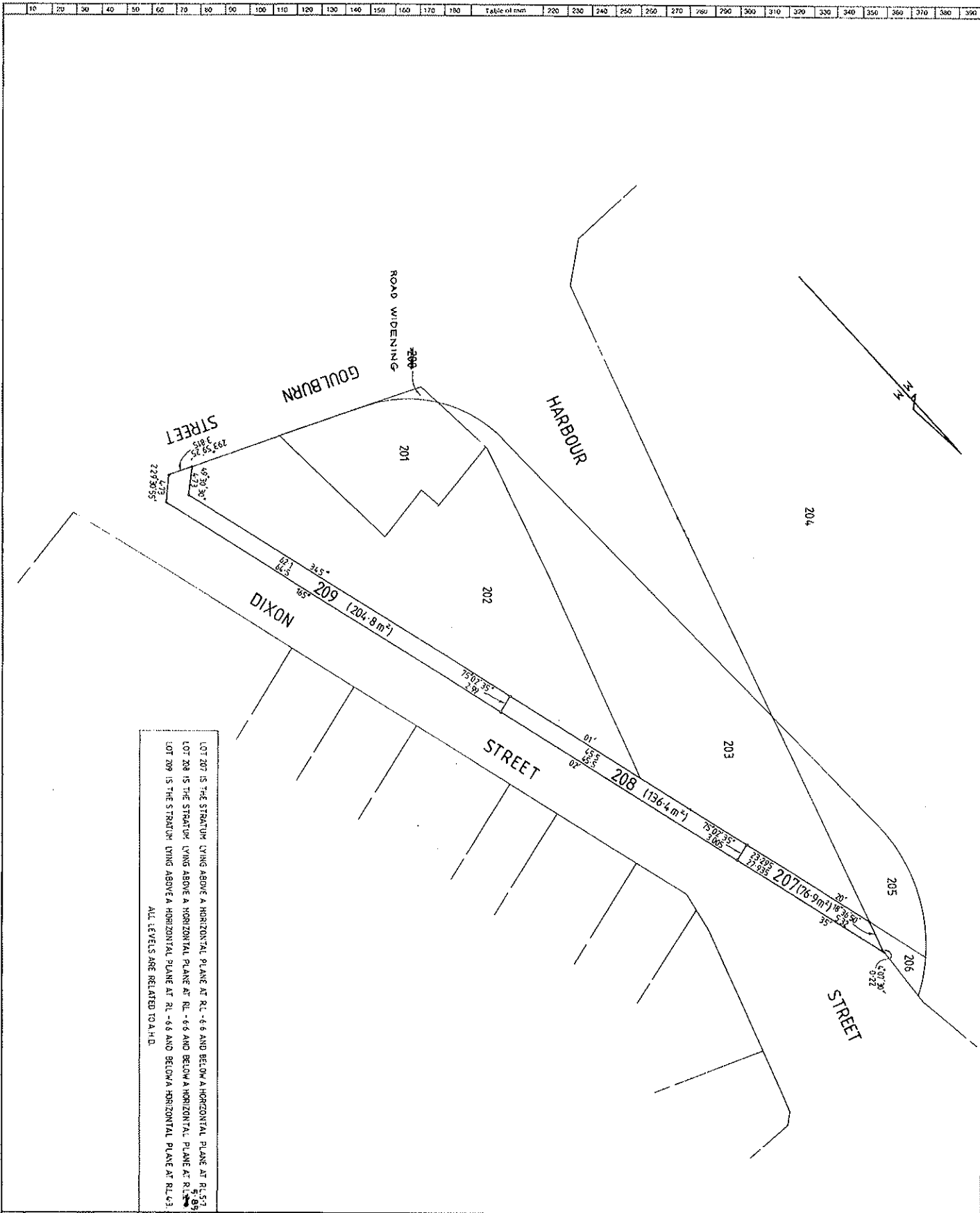
SURVEYORS REFERENCE: SD 0812



Plan Drawing only to appear in this space

SD 0715/23 SURVEYORS REFERENCE

DESIGNATION Y ADDED IN LTO ON 2-7-77



LOT 201 IS THE STRATUM LYING ABOVE A HORIZONTAL PLANE AT RL -6.6 AND BELOW A HORIZONTAL PLANE AT RL -5.2
 LOT 202 IS THE STRATUM LYING ABOVE A HORIZONTAL PLANE AT RL -6.6 AND BELOW A HORIZONTAL PLANE AT RL -5.2
 LOT 203 IS THE STRATUM LYING ABOVE A HORIZONTAL PLANE AT RL -6.6 AND BELOW A HORIZONTAL PLANE AT RL -5.2
 LOT 204 IS THE STRATUM LYING ABOVE A HORIZONTAL PLANE AT RL -6.6 AND BELOW A HORIZONTAL PLANE AT RL -5.2
 LOT 205 IS THE STRATUM LYING ABOVE A HORIZONTAL PLANE AT RL -6.6 AND BELOW A HORIZONTAL PLANE AT RL -5.2
 LOT 206 IS THE STRATUM LYING ABOVE A HORIZONTAL PLANE AT RL -6.6 AND BELOW A HORIZONTAL PLANE AT RL -5.2
 ALL LEVELS ARE RELATED TO A.H.D.

Plan Drawing only to appear in this space

Surveyor's Reference: SD0715/83

DP 771840 Registered 10.2.1998 This is sheet 3 of my plan of 3 sheets dated 131 AUGUST 1987 Surveyor registered under Surveyors Act 1929 <i>Abraham</i> This is a true and correct copy of the original plan signed and covered by my Certificate No. 11	Council/Chair This plan was made a rule of court in my name on Plan Form 2
---	---

Reduction Ratio 1:400

PLAN FORM 2

Standard and JCRS only

Plan Drawing only to appear in this space

OFFICE USE ONLY

DP 739093

Registered 10/2/1988

C.A.

THE SYDNEY TORRENS & O.O.
SYSTEM
SUBDIVISION
PA 17081 SUBDIVISION
BUT. NEAR PARISH, CITY OF SYDNEY
SH 101

LET. PLAN SEE SCHEDULE 6000

PLAN
OF SUBDIVISION OF
DARLING HARBOUR LANDS

Reduction Scale 1:1000



City
SYDNEY

Location
DARLING HARBOUR

Parish
ST ANDREW

County
CUMBERLAND

This is a plan of the land shown in the plan.

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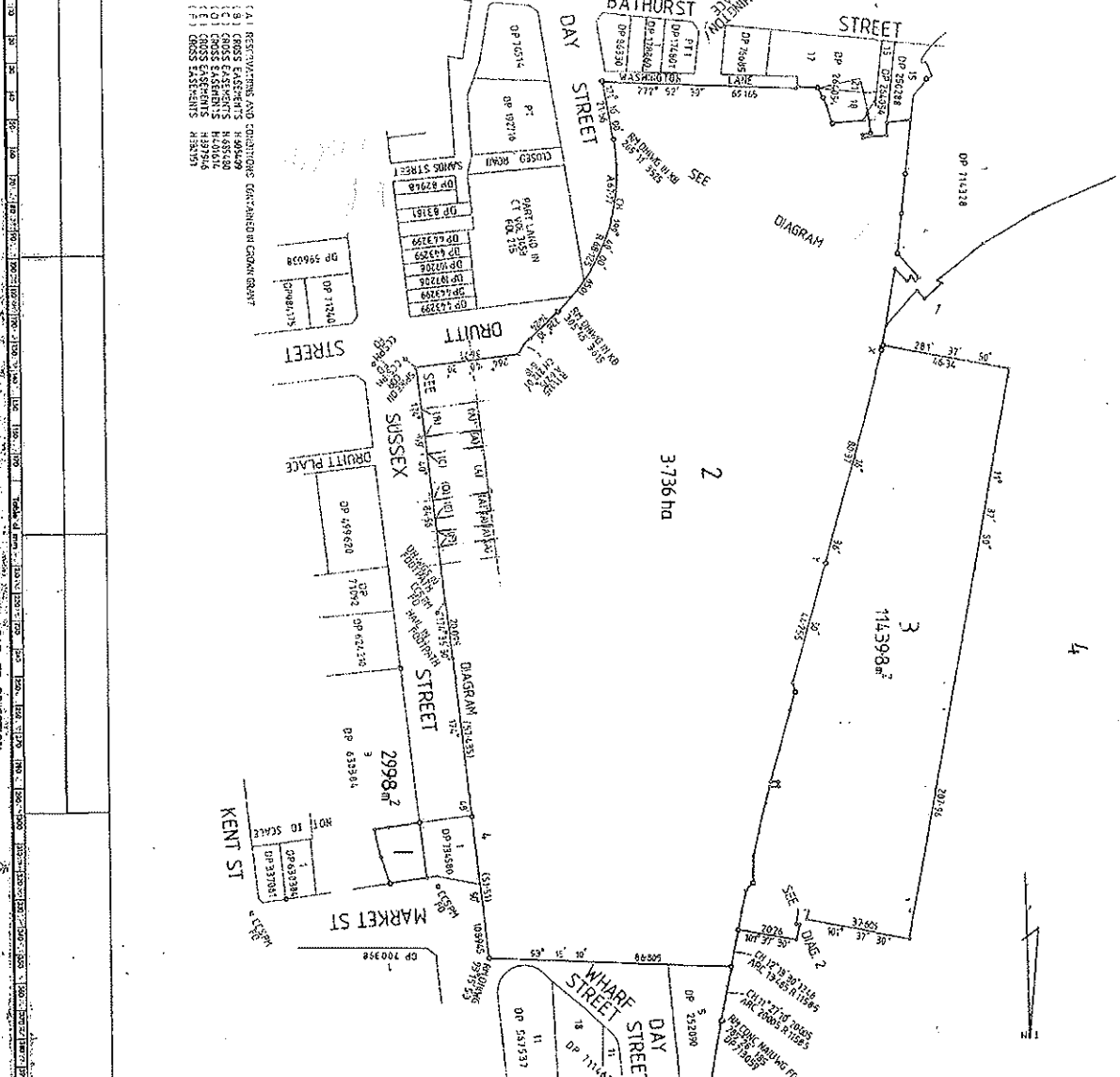
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- (A) RESERVATIONS AND CONDITIONS CONTAINED IN COVENANT
- (B) CROSS EASEMENTS
- (C) CROSS EASEMENTS
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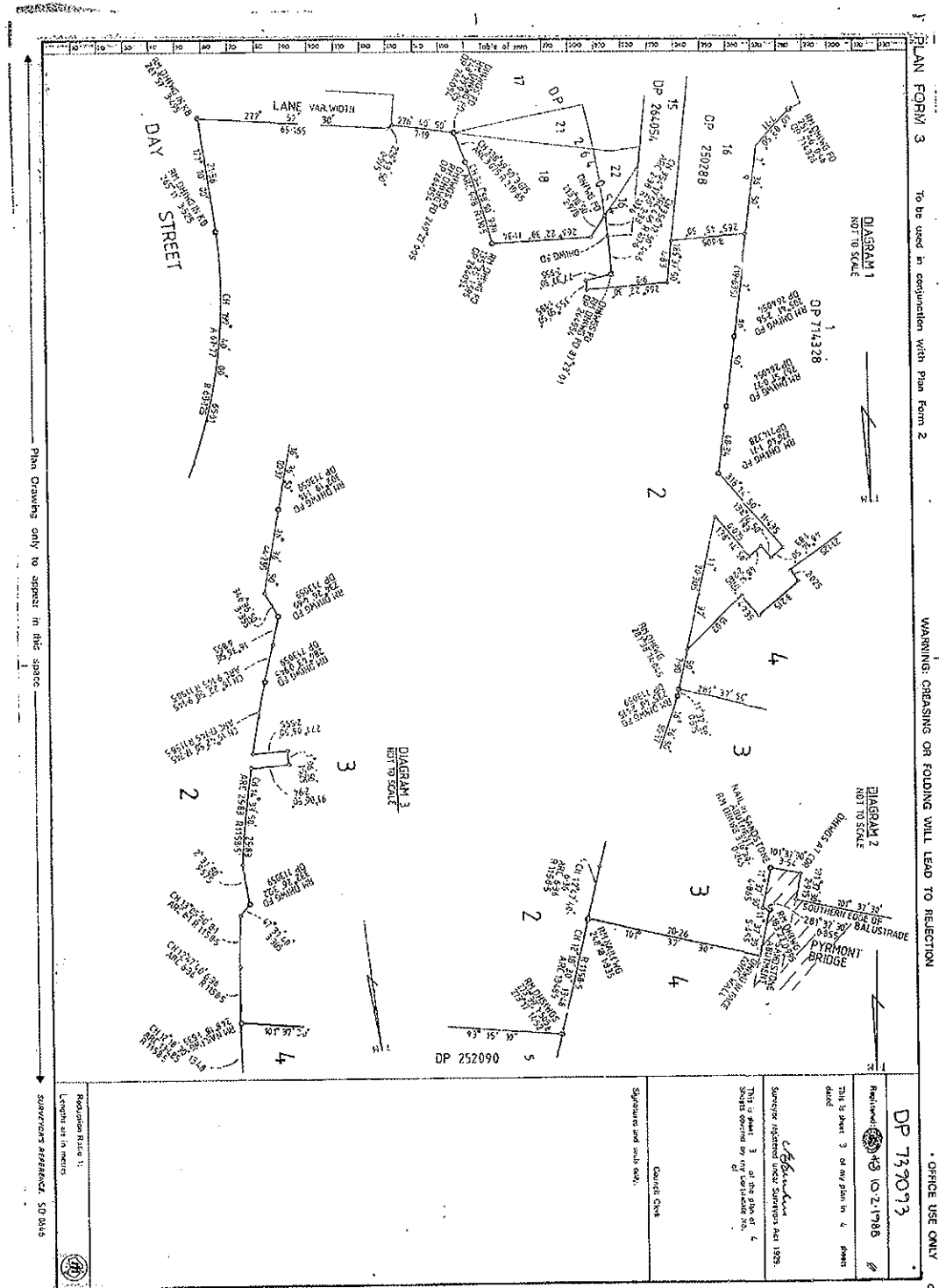
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WARNING: CREASING, OR FOLDING WILL LEAD TO REJECTION

This negative is a photograph made as a permanent record of a document in the custody of the Registrar General this day.

18th February, 1988

[illegible]



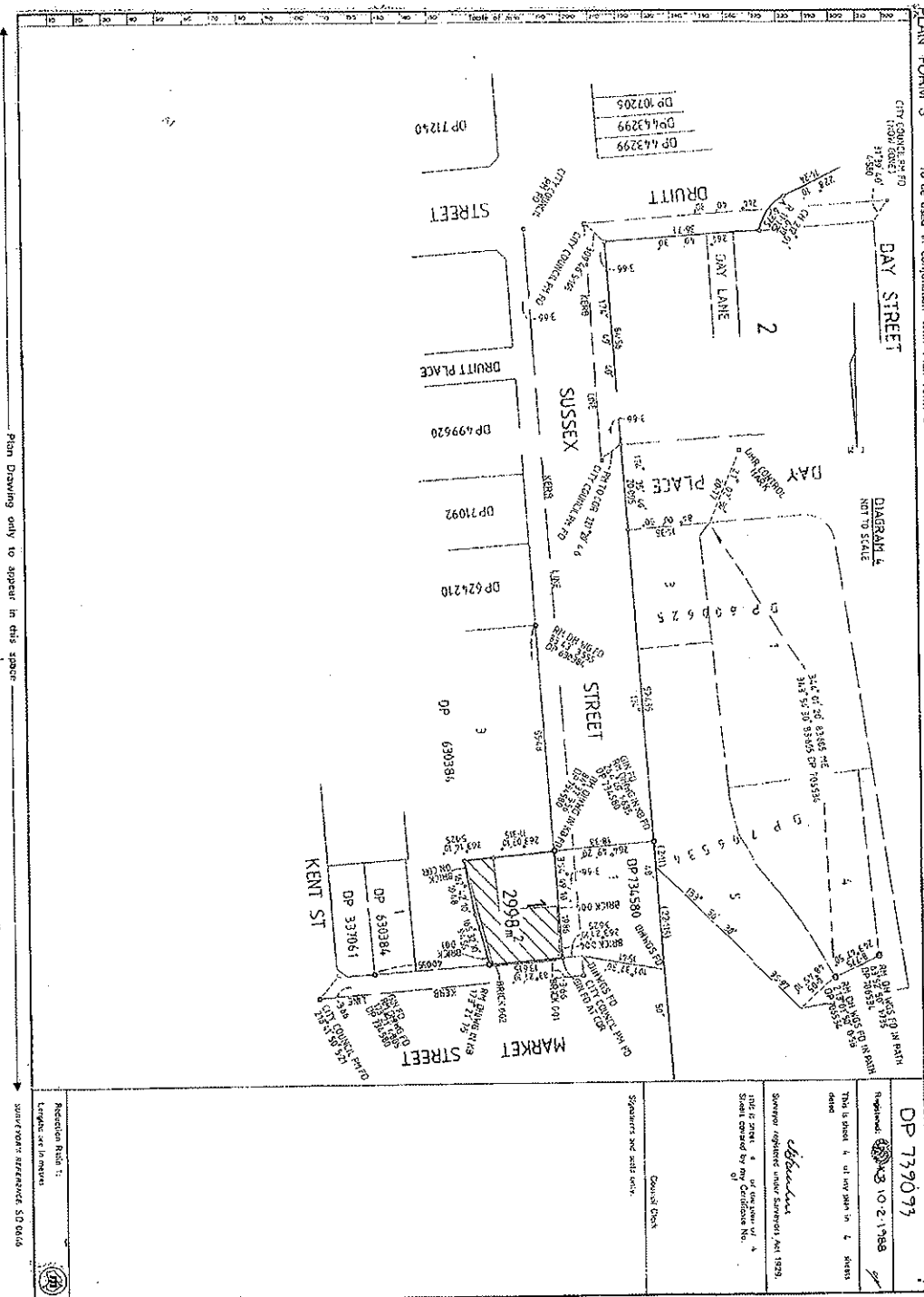
This negative is a photograph made as a permanent record of a document in the custody of the Registrar General this day.

18th February, 1988

PLAN FORM 3 To be used in conjunction with Plan Form 2

WARNING: CREASING OR FOLDING WILL LEAD TO REJECTION

OFFICE USE ONLY



DP 739073

Registered 10-2-1988

This is sheet 4 of any part in 2 sheets

Surveyor's signature (not necessary after 1978)

Site is shown 4 of the plan of 4

Sheet covered by any Certificate No.

Surveyor's name and address

Surveyor's name and address

Surveyor's name and address

Surveyor's name and address

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TO 20 30 40 50 60 70 80 90 100 110 120 130 140

This negative is a photograph made as a permanent record of a document in the custody of the Registrar General this day 18th February, 1988



TH 27 MAR 2007

Our Ref: D07/028097
Your Ref: E21073F

23 March 2007

Attention: Mr Todd Hore
EIS
PO Box 976
NORTH RYDE BC NSW 1670

Dear Mr Hore

RE SITE: Lot 318 DP 871455, Darling Walk, Darling Harbour

I refer to your search request of 22 March 2007 requesting information on licences to Keep Dangerous Goods for the above site.

A search of the Stored Chemical Information Database (SCID) and the microfiche records held by WorkCover has not located any records pertaining to the above-mentioned premises.

If you have any further queries, please contact Dangerous Goods Licensing staff on (02) 4321 5500.



Ian Gough
Team Leader
Dangerous Goods

WorkCover. **Watching out for you.**

APPENDIX D

SOIL AND GROUNDWATER SAMPLING PROTOCOLS

These protocols specify the basic procedures to be used when sampling soils or groundwater for environmental site assessments undertaken by Environmental Investigation Services. The purpose of these protocols is to provide standard methods for: sampling, decontamination procedures for sampling equipment, sample preservation, sample storage and sample handling. Deviations from these procedures must be recorded.

SOIL SAMPLING

- (i) prepare a test pit/borehole log.
- (ii) Layout sampling equipment on clean plastic sheeting to prevent direct contact with ground surface. The work area should be at a distance from the drill/rig excavator such that the drill rig/excavator can operate in a safe manner.
- (iii) Ensure all sampling equipment has been decontaminated prior to use.
- (iv) Remove any surface debris from the immediate area of the sampling location.
- (v) Collect samples and place in a glass jar with a Teflon sea. This should be undertaken as quickly as possible to prevent the loss of volatiles. If possible, fill the glass jars completely.
- (vi) Label the jar with the EIS job number, sample location (eg. TP1), sampling interval and date. If more than one sample container is used, this should also be indicated (eg. 2 = Sample jar 1 of 2 jars).
- (vii) Photoionisation detector (PID) screening of volatile organic compounds (VOCs) should be undertaken on samples using the soil sample headspace method. Headspace measurements are taken following equilibration of the headspace gasses in partly filled glass jars. PID headspace data is recorded on the borehole/test pit log and the chain of custody forms.
- (viii) Record the lithology of the sample and sample depth on the borehole/test pit log in accordance with AS1726-1993.
- (ix) Store the sample in a sample container cooled with ice or chill packs. On completion of the sampling the sample container should be delivered to the lab immediately or stored in the refrigerator prior to delivery to the lab.
- (x) Check for the presence of groundwater after completion of each borehole using an electronic dip metre or water whistle. Boreholes should be left open until the end of fieldwork. All groundwater levels in the boreholes should be rechecked on the completion of the fieldwork.
- (xi) Backfill the boreholes/test pits with the excavation cuttings or clean sand prior to leaving the site.

DECONTAMINATION PROCEDURES FOR SOIL SAMPLING EQUIPMENT

- (i) All of the equipment associated with the soil sampling procedure should be decontaminated between every sampling location.
- (ii) The following equipment and materials are required for the decontamination procedure:
 - Phosphate free detergent (Extran 100)
 - Tap water
 - Two buckets
 - Stiff brushes
 - Plastic sheets
- (iii) Ensure the decontamination materials are clean prior to proceeding with the decontamination.
- (iv) Fill both buckets with clean tap water and add phosphate free detergent to one bucket.
- (v) In the bucket containing the detergent scrub the sampling equipment until all the material attached to the equipment has been removed.

- (vi) Rinse sampling equipment in the bucket containing tap water.
- (vii) Place cleaned equipment on clean plastic sheets.

If all materials are not removed by this procedure, high-pressure water cleaning is recommended. If any equipment is not completely decontaminated by both these processes that equipment should not be used until it has been thoroughly cleaned.

GROUNDWATER SAMPLING

Groundwater samples are more sensitive to contamination than soil samples and therefore adhesion to this protocol is particularly important to obtain reliable, reproducible results. The recommendations details in AS2306.1 are considered to form a minimum standard.

The basis of this protocol is to maintain the security of the borehole and obtain accurate and representative groundwater samples. The following procedure should be used for collection of groundwater samples from previously installed piezometers.

- (i) After piezometer installation, at least four bore volumes should be pumped from the piezometers to remove any water introduced during the drilling process. Piezometers should then be left to recharge for at least five days before purging and sampling. Prior to purging or sampling the condition of each well should be observed and any anomalies recorded on the field data sheets. The following information should be noted: the condition of the well, noting any signs of damage, tampering or complete destruction; the condition and operation of the well lock; the condition of the protective casing and the cement footing (raised or cracked); and, the presence of water between protective casing and well.
- (ii) Take the groundwater level from the collar of the piezometer using an electronic dipmeter. The collar level should be taken during the site visit using a dumpy level and staff.
- (iii) Purging and sampling of piezometers should generally be done on the same site visit. Layout and organize all equipment

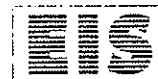
associated with groundwater sampling in a location where they will not interfere with the sampling procedure and will not pose a risk of contaminating samples. Equipment generally required includes:

- New disposable polyethylene bailer and sufficient cord OR submersible pump.
 - Micropore filtration system (for heavy metals samples).
 - Filter paper (glass fibre and 0.45µm).
 - Buckets with volume increments.
 - Sample containers – at least 1 x Teflon bottle with 1ml nitric acid, 1 x 75mL glass vial and 2 x 1L amber glass bottles for each piezometer.
 - pH/Cond/Eh/T meters.
 - Glass jars for purged samples.
 - Esky and ice.
 - Latex gloves.
 - Distilled water (for cleaning).
 - Electronic dipmeter.
 - Groundwater sampling forms and notebook.
 - Aluminium foil and labels.
- (iv) Clean the Micropore filtration system thoroughly with distilled water prior to use and between each sample. Filter paper should be changed between samples. 0.45µm filter paper should be placed below the glass fibre filter paper in the filtration system.
 - (v) Ensure all non-disposable sampling equipment is decontaminated or that new disposable equipment is available prior to any work commencing at a new location. The procedure for decontamination of groundwater equipment is outlined at the end of this section.
 - (vi) Disposable gloves should be used whenever samples are taken to protect the sampler and to assist in avoidance of contamination.

- (vii) Purge at least four bore volumes from the well. Take pH, conductivity, redox potential, and temperature measurements of the purged groundwater at regular intervals during purging. (Say, every 5-10 litres if abundant groundwater and every 1 litre if only limited groundwater is encountered). Groundwater condition measurements should be taken from a sample in a clean glass jar which has been taken directly from the sampling equipment (either pump or bailer). Electrodes should be placed in the sample after the electrodes have been rinsed with distilled water. Purged volumes and groundwater measurements should be recorded on the field sampling sheet. An assessment of the turbidity of the sample should also be made based on three categories: silty, opaque and clear.
- (viii) Prepare all sample bottles. Label bottles with EIS job number, borehole number and date of collection.
- (ix) Fill amber sample bottles and BTEX vial directly from pump or bailer. Ensure sampling equipment does not touch sample containers. Sample bottles and vials must be filled to the brim, so that a reverse meniscus is formed, seal with aluminium foil and then cap. Check that no air has entered the sample invert and check for bubbles.
- (x) Fill vacuum filtration system and turn on filter pump.
- (xi) Undertake pH/Cond/Eh/T of a sample taken in a clean glass jar used only for groundwater condition measurements. Turn the meters on and insert the electrodes into the sample. Record the measurements when the instruments have stabilized, then discard the sample. Clean the electrodes with distilled water between measurements.
- (xii) When the sample filtering is complete (note: at least 50mL of filtered sample is required for heavy metal analysis), decant the filtered sample into a Teflon bottle containing nitric acid. Check label of sample bottle to ensure container has been treated with nitric acid and not sulfuric acid. Clean the filtration system with distilled water and replace the filters ready for the next sample.
- (xiii) Photoionisation detector (PID) screening of volatile organic compounds (VOC) should be undertaken on groundwater samples using the sample headspace method during fieldwork. VOC data is obtained from partly filled glass jar samples following equilibration of the headspace gases. The PID headspace data should be included on the chain of custody forms and borehole logs.
- (xiv) Store the sample in a sample container cooled with ice or chill packs. On completion of the sampling the sample container should be delivered to the lab immediately or stored in the refrigerator prior to delivery to the lab.
- (xv) Record the sample on the appropriate log in accordance with AS1726-1993. At the end of each water sampling complete a chain of custody form.

DECONTAMINATION PROCEDURE FOR GROUNDWATER SAMPLING EQUIPMENT

- (i) All of the equipment associated with the groundwater sampling procedure should be decontaminated between every sampling location.
- (ii) The following equipment and materials are required for the decontamination procedure:
 - Phosphate free detergent (Extran 100).
 - Tap water.
 - Distilled water.
 - Two buckets.
 - Plastic sheets.



- (iii) Fill one bucket with clean tap water and phosphate free detergent, and one bucket with distilled water.
- (iv) Flush tap water and detergent through pump. Wash sampling equipment and pump head using brushes in the bucket containing detergent until all materials attached to the equipment are removed.
- (v) Flush pump with distilled water.

- (vi) Change water and detergent solution after each sampling location.
- (vii) Rinse sampling equipment in the bucket containing distilled water.
- (viii) Place cleaned equipment on clean plastic sheets.

If all materials are not removed by this procedure that equipment should not be used until it has been thoroughly cleaned.



QA/QC DEFINITIONS

The QA/QC terms used in this report are defined below. The definitions are in accordance with current US EPA SW-846 (1994) methods and those described in Environmental Sampling and Analysis, A Practical Guide, (H. Keith 1991).

Practical Quantitation Limit (PQL), Limit of Reporting (LOR) and Estimated Quantitation Limit (EQL)

These terms all refer to the concentration above which results can be expressed with a minimum 95% confidence level. The laboratory reporting limits are generally set at ten times the standard deviation for the Method Detection limit (MDL) for each specific analyte. For the purposes of this report the LOR, PQL, and EQL are considered to be equivalent.

When assessing laboratory data it should be borne in mind that values at or near the PQL have two important limitations.

"The uncertainty of the measurement value can approach, and even equal, the reported value. Secondly, confirmation of the analytes reported is virtually impossible unless identification uses highly selective methods. These issues diminish when reliably measurable amounts of analytes are present. Accordingly, legal and regulatory actions should be limited to data at or above the reliable detection limit", Keith (1991).

Accuracy

The proximity of an averaged result to the true value, where all random errors have been statistically removed. Accuracy is measured by percent recovery. Acceptable limits for accuracy generally lie between 70% to 130% recoveries. Certain laboratory methods may allow for values that lie outside these limits.

Precision

The degree to which data generated from repeated measurements differ from one another due to random errors. Precision is measured using the standard deviation or Relative Percent Difference (RPD). Acceptable targets for precision in this report will be less than 50% RPD for concentrations greater than ten times the PQL, less than 75% RPD for concentrations between five and ten times the PQL and less than 100% RPD for concentrations that are less than five times the PQL.

Blanks

The purpose of laboratory and field blanks is to check for artifacts and interferences that may arise during sampling and analysis.

Matrix Spikes

Samples are spiked with laboratory grade standards to detect interactive effects between the sample matrix and the analytes being measured. Matrix Spikes are reported as a percent recovery and are prepared for 1 in every 20 samples. Sample batches that contain less than 20 samples



may be reported with a Matrix Spike from another batch. The percent recovery is calculated using the formula;

$$\frac{(\text{spiked sample result} - \text{sample result})}{\text{concentration of spike added}} \times 100$$

Acceptable recovery limits are 70% to 130%.

Surrogate Spikes

Samples are spiked with a known concentration of compounds that are chemically related to the analyte being investigated but unlikely to be detected in the environment. The purpose of the Surrogate Spikes is to check the accuracy of the analytical technique. Surrogate Spikes are reported as percent recovery.

Duplicates

Laboratory duplicates measure precision, expressed as Relative Percent Difference. Duplicates are prepared from a single field sample and analysed as two separate extraction procedures in the laboratory. The RPD is calculated using the formula:

$$\frac{|D1 - D2|}{|(D1 + D2)/2|} \times 100$$

where D1 is the sample concentration and D2 is the duplicate sample concentration.